

## Cost-efficient cultivation of *Spirulina platensis* by chemical absorption of CO<sub>2</sub> into medium containing NaOH

Joo-Young Jung\*, Ji-Won Yang\*, Kyochan Kim\*\*, Kwon-Tack Hwang\*\*\*,  
Simon MoonGeun, Jung\*\*\*\*,†, and Jong-Hee Kwon\*\*\*\*\*,†

\*Advanced Biomass R&D Center, Building W1-3, KAIST, 291, Daehak-ro, Yuseong-gu, Daejeon 305-701, Korea

\*\*Department of Chemical and Biomolecular Engineering, KAIST, 291, Daehak-ro, Yuseong-gu, Daejeon 305-701, Korea

\*\*\*Department of Food and Nutrition, Nambu University, Gwangju 506-706, Korea

\*\*\*\*SK Innovation Global Technology, 325, Expo-ro, Yuseong-gu, Daejeon 305-712, Korea

\*\*\*\*\*Department of Food Science & Technology and Institute of Agriculture & Life Science,  
Gyeongsang National University, Jinju 660-701, Korea

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**Abstract**—*Spirulina* is one of the promising photosynthetic microorganisms as the source for food, cosmetic, and other value-added products such as phycocyanin. However, its production cost associated with cultivation is expensive because of high concentration of bicarbonate (76 wt%) in standard Zarrouk medium for *Spirulina*. Bicarbonate not only acts as a carbon source but also helps maintaining culture medium in alkaline condition, which is essential for growth of *Spirulina*. The present study demonstrates that bicarbonate (16.8 g/L) in standard Zarrouk medium was completely replaceable by chemical CO<sub>2</sub> absorption using 0.2 M NaOH and 154.2 mmol CO<sub>2</sub> gas could be absorbed in the form of NaHCO<sub>3</sub>/Na<sub>2</sub>CO<sub>3</sub> into 1 L medium during the process. This process was incorporated into *Spirulina* cultivation, which enabled us to reduce the total cost of preparation of Zarrouk medium by 34.3% without sacrificing biomass and pigments production. In addition, another important use of NaOH is achievement of cost-efficient and eco-friendly sanitization of culture medium.

Keywords: Chemical Absorption, Sodium Hydroxide, Carbon Dioxide, Zarrouk Medium, *Spirulina platensis*

### INTRODUCTION

*Spirulina platensis* (hereafter *S. platensis*) is a blue-green algae with many beneficial properties. Used as a food source for centuries in Africa, America, and Asia, *S. platensis* has high quality proteins, pigments, carbohydrates, lipids, and many other essential nutrients such as vitamin, anti-oxidants [1]. Although mass production of *S. platensis* has been in progress, due to high usage of bicarbonate in growth medium for *S. platensis*, its production cost associated with cultivation is still relatively expensive. Bicarbonate is not only used as a carbon source for triacylglycerol and starch but also helps to maintain alkaline conditions, which are essential for the growth of *Spirulina* [2].

Carbon dioxide (CO<sub>2</sub>), which constitutes up to 68% of total emissions of greenhouse gases (GHGs), has been an important issue in global economics and politics, as well as the environment [3,4]. Absorption is a common physical or chemical process for the treatment of industrial gas streams. Inorganic bicarbonate for cultivation of microalgae can be supplied by chemical absorption of gaseous CO<sub>2</sub>, resulting in soluble inorganic carbon such as NaHCO<sub>3</sub> and Na<sub>2</sub>CO<sub>3</sub> [5-7]. Among the chemicals used for CO<sub>2</sub> absorption, strong hydroxides such as sodium hydroxide have the advantages of high

load capacity and fast reaction time due to high binding energy of chemical sorbents [6].

*S. platensis* is in general cultivated in Zarrouk medium, which contains sodium bicarbonate (76 wt%) as a major component (Zarrouk, 1966). The potential use of absorption of CO<sub>2</sub> using NaOH was investigated and presented in this research, in terms of chemical sterilization and replacement of NaHCO<sub>3</sub> by NaOH in standard Zarrouk medium for cultivation of *S. platensis*. The overall process is illustrated in Fig. 1. The production of biomass and photosynthetic pigments such as chlorophyll and phycocyanin was measured to validate sustainable growth of *S. platensis* in the prepared Zarrouk medium via chemical absorption of CO<sub>2</sub> using NaOH.

### MATERIAL AND METHODS

#### 1. Preparation of Cultivation Medium and Chemical Absorption of CO<sub>2</sub> by 0.2 M NaOH

*S. platensis* was obtained from the Korea Marine Microalgae Culture Center (KMMCC). Standard Zarrouk medium consists of 16.8 g/L NaHCO<sub>3</sub>, 0.5 g/L K<sub>2</sub>HPO<sub>4</sub>, 2.5 g/L NaNO<sub>3</sub>, 1.0 g/L K<sub>2</sub>SO<sub>4</sub>, 1.0 g/L NaCl, 0.2 g/L MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.04 g/L CaCl<sub>2</sub>·2H<sub>2</sub>O, 0.01 g/L FeSO<sub>4</sub>·7H<sub>2</sub>O, 0.08 g/L EDTA, and 1 ml micronutrient solution. The micronutrient solution consisted of 2.86 g/L H<sub>3</sub>BO<sub>3</sub>, 1.81 g/L MnCl<sub>2</sub>·4H<sub>2</sub>O, 0.222 g/L ZnSO<sub>4</sub>·4H<sub>2</sub>O, 0.0177 g/L Na<sub>2</sub>MoO<sub>4</sub>, and 0.079 g/L CuSO<sub>4</sub>·5H<sub>2</sub>O (Zarrouk et al. 1966). For chemical absorption of CO<sub>2</sub>, 0.2 M NaOH was added to the modified Zarrouk medium, lacking sodium

†To whom correspondence should be addressed.

E-mail: jungmoongeun@sk.com, jhkwon@gnu.ac.kr

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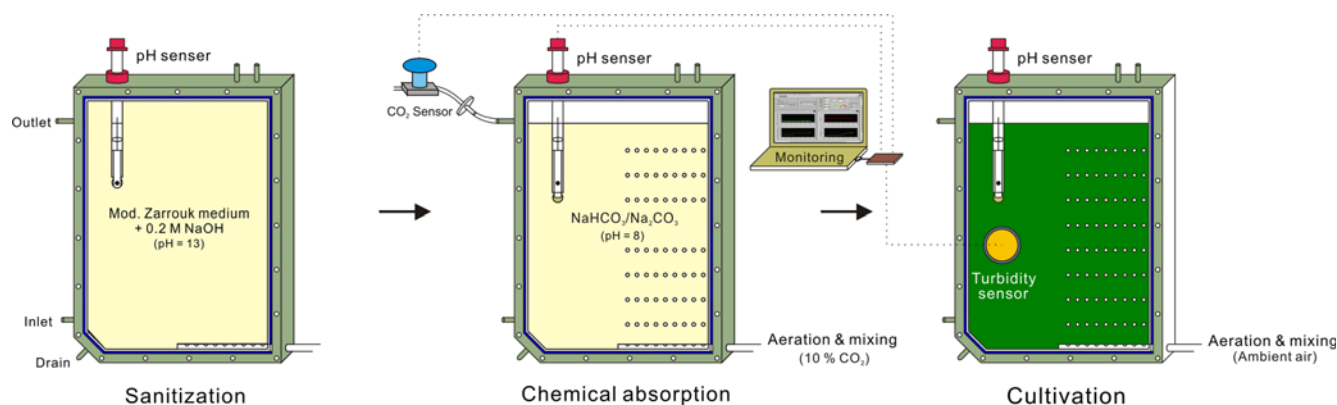


Fig. 1. Overall process diagram of chemical absorption by NaOH solution from sanitization to cultivation.

bicarbonate. The concentration of NaOH was set to match the amount of bicarbonate in standard Zarrouk medium. This modified medium was added to a closed 2 L flat-panel photobioreactor system, followed by aeration with a constant flow of  $200 \text{ ml min}^{-1}$  air enriched with 10%  $\text{CO}_2$  (v/v) from a perforated plastic tube embedded in the bottom of the reactor. Two modified Zarrouk media at pH 8 and 9 were set by time-dependent  $\text{CO}_2$  aeration, i.e., 10 and 7.5 hours, respectively, while standard Zarrouk medium has a pH of 8.7. During cultivation of *S. platensis*, temperature was maintained at  $25^\circ\text{C}$ , and culture was continuously illuminated at a light intensity of  $100 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$  and mixed with ambient air containing 0.04%  $\text{CO}_2$  at a constant flow of  $200 \text{ ml min}^{-1}$ .

## 2. Sanitization by NaOH

Prior to chemical absorption of  $\text{CO}_2$ , the reactor system was chemically sterilized by incubation with 0.2 M NaOH solution (pH=13), while standard Zarrouk medium was sterilized in an autoclave. Disinfectant effect of 0.2 M NaOH as a function of incubation time was verified by CFU tests ( $n=10$ ) using two different agars, TYG (5 g/L tryptone, 5 g/L yeast extract, 1 g/L glucose, 1 g/L  $\text{K}_2\text{HPO}_4$ , 15 g/L Bacto™ agar) and LB (25 g/L LB Broth, 15 g/L Bacto™ agar). Bacteria-contaminated medium was treated with 0.2 M NaOH for different incubation time: 1, 2, 3, and 4 hours. Medium treated with 0.2 M NaOH was plated onto the agars and incubated for 2 days at  $37^\circ\text{C}$  under light intensity of  $50 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ . After this short incubation period, the number of individual bacterial colonies on each plate was counted. Bacterial populations were identified by DNA sequencing based on 16S rDNA blast library.

## 3. Measurement of pH and $\text{CO}_2$ Concentration

$\text{CO}_2$  concentration absorbed in culture medium was calculated in terms of the detected partial pressure of  $\text{CO}_2$  in emission gas by a BCP- $\text{CO}_2$  sensor (BlueSens GmbH, Herten, Germany). The sensor was directly connected with the exhaust-gas stream of the fermenter using a flow adapter. The gas flow rate was estimated as  $200 \text{ mL min}^{-1}$  to calculate a molar concentration of  $\text{CO}_2$  in exhaust gas. The change of pH level was on-line monitored by a pH meter (Horiba Co. Ltd., Kyoto, Japan) equipped inside the photobioreactor. Measurements of pH value and  $\text{CO}_2$  concentration continued until both reached a steady state.

## 4. Analytical Methods

To measure biomass concentration, equal volumes of biomass

culture were filtered through GF/C glass fiber filter paper, which was pre-dried at  $50^\circ\text{C}$  for 24 h. The differences between weights of filter paper before filtration and dried filter paper after filtration were calculated. Concentrations of chlorophyll *a* [8] and phycocyanin [9] were measured during cultivation.

## RESULTS

### 1. Sanitization by NaOH

The antimicrobial activity of aqueous solutions of 0.2 M NaOH was investigated by CFU tests on LB and TYG agar plates. Medium contaminated by bacterial population indicated an initial colony-forming unit of  $1.57 \pm 0.11 \times 10^5 \text{ CFU mL}^{-1}$  on LB agar plates and  $1.24 \pm 0.14 \times 10^5 \text{ CFU mL}^{-1}$  on TYG agar plates. In contaminated medium, there were the majority of five bacterial strains detected; *Tristrella*, *Pseudomonas*, *Stappia*, *Labrenzia* and *Microbacterium*. After 1 hour incubation with 0.2 M NaOH solution, no colony was formed on either plate.

### 2. Chemical Absorption of $\text{CO}_2$ by Aqueous NaOH Solution in Modified Medium

Fig. 2 shows pH changes of medium and the concentration of  $\text{CO}_2$  in the emitted gas over 12 hours, with both appearing as double logistic sigmoid curves. More than 95% of the infused  $\text{CO}_2$  was converted to aqueous form of carbon dioxide for the first 2 hours, and half of the total infused  $\text{CO}_2$  was still absorbed until the first sigmoid change appeared.

The pH value of the solution is of major importance in equilibrium of this reaction system. Different equilibrium conditions were observed following absorption of  $\text{CO}_2$  gas by alkaline aqueous solution. The pH value decreased nonlinearly as a function of treatment time, and its value was not lower than pH 8.0 due to the formation of a carbonate-bicarbonate buffer (Fig. 2(a)). The fraction of bicarbonate among inorganic carbon sources decreased when the pH decreased. After 10 hours of  $\text{CO}_2$  injection, the solution was completely saturated with aeration containing 9.87% of  $\text{CO}_2$  (Fig. 2(a)).

Fig. 2(b) presents the amount of  $\text{CO}_2$  absorbed into modified medium when aerated with 10 %  $\text{CO}_2$  gas at a rate of  $200 \text{ ml min}^{-1}$ . While the pH decreased from 13 to 8,  $154.2 \text{ mmol CO}_2$  gas was absorbed into 1 L of medium. Assuming working volume of 1,000 L for the preparation of modified medium, this process could cap-

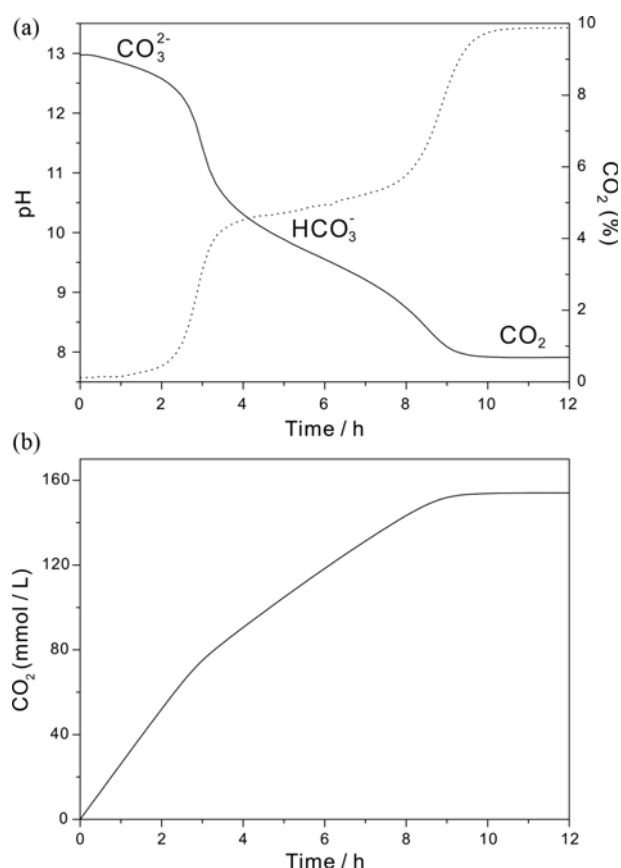


Fig. 2. (a) Chemical absorption of 10% (v/v) CO<sub>2</sub> gas into modified Zarrouk medium containing 0.2 M sodium hydroxide. Changes in pH value inside the reactor are indicated as solid lines. CO<sub>2</sub> concentration in the gas emitted from the reactor is recorded as dashed lines. (b) Total absorbed inorganic carbon concentration in medium.

ture 6.78 kg of CO<sub>2</sub> gas prior to cultivation of *S. platensis*. In addition, the emitted CO<sub>2</sub> gas into air could be recycled and reused for additional chemical absorption process.

### 3. Growth of *Spirulina platensis*

Cultivation conditions such as pH, temperature and nutrients have an effect on biomass accumulation by *S. platensis*; 1.5–3 g/L biomass accumulation by *S. platensis* was reported in previous research [10,11]. In this research, 2.5 g/L of biomass was accumulated in photoautotrophic conditions. From the cultivation results, no significant change in biomass and pigment production was monitored among cells grown in the media prepared by chemical absorption and in the standard Zarrouk medium (Fig. 3). In conclusion, this research demonstrated that bicarbonate, essential for growth of *S. platensis*, could be successfully replaced by chemical absorption of CO<sub>2</sub> using 0.2 M NaOH without loss in production of biomass and pigments.

## DISCUSSION

Post-combustion CO<sub>2</sub> capture can be accomplished by chemical absorption, adsorption, membrane separation, cryogenic distillation, and chemical looping combustion [12–14]. Although these

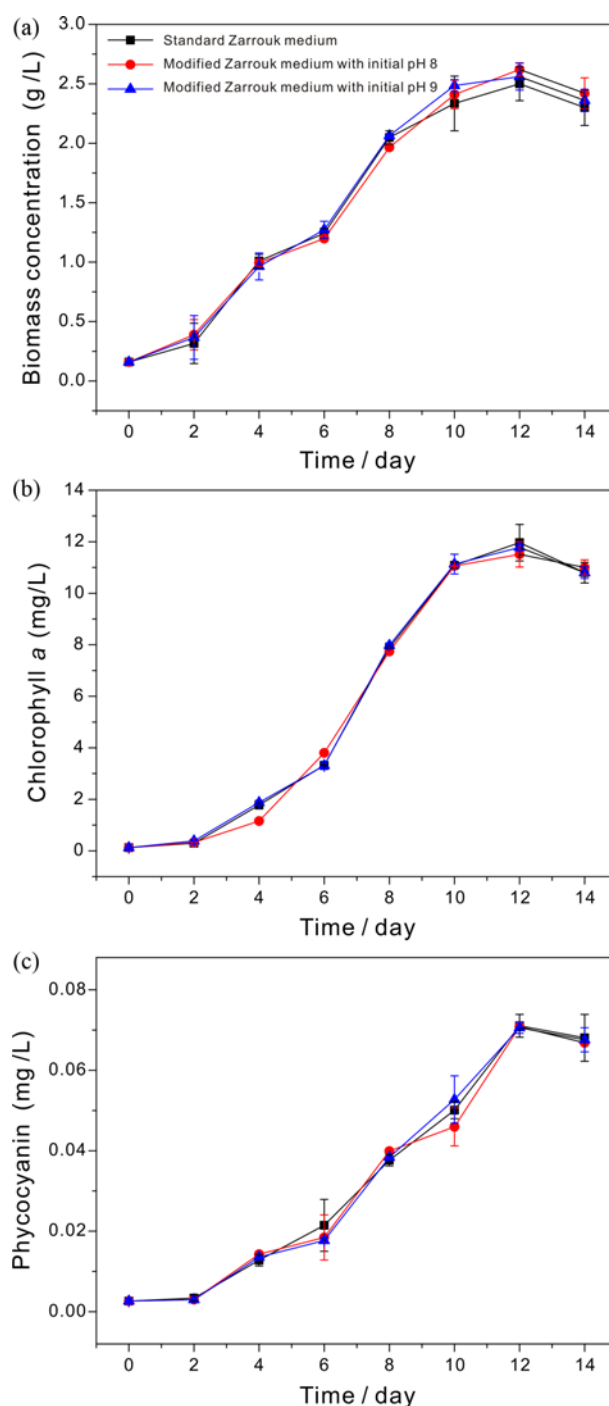


Fig. 3. Biomass (a), C-phycoerythrin (b), and chlorophyll a (c) contents of *S. platensis* cells grown in modified Zarrouk media (pH 8 and 9) and standard Zarrouk medium. The plotted values indicate the average value of three independent measurements within standard errors.

physical and chemical methods are well-developed and show high efficiency and capacity for CO<sub>2</sub> capture, they have a few drawbacks in high energy consumption, large space requirements, and difficulties in utilizing the captured CO<sub>2</sub> for valuable products [15]. Moreover, additional energy penalties can be incurred from compression and disposal of concentrated CO<sub>2</sub> complexes in these pro-

cesses [16]. Conventional methods such as underground CO<sub>2</sub> storage, fertilizer production, and usage in food and beverage industry are limited in their ability to store all CO<sub>2</sub> captured from fossil fuel power plants [17]. New alternatives of CO<sub>2</sub> capture technology, including biological fixation using microalgae and cyanobacteria, have recently received attention for their ability to biochemically convert CO<sub>2</sub> into biomass, which can be utilized as a platform for the production of biofuels, foods, cosmetics, and medications [18]. However, biological CO<sub>2</sub> fixation rate by microalgae is much lower than those of physical and chemical technologies of CO<sub>2</sub> capture. Furthermore, additional methods are required to drastically reduce the amount of CO<sub>2</sub> lost to the atmosphere due to its low solubility in pure water.

These limitations of respective treatment of CO<sub>2</sub> can be overcome by a method involving combinations of biological methods and chemical absorption. NaOH present in medium will react with dissolved CO<sub>2</sub> gas, to the extent determined by the pH of culture medium. This will cause consumption of OH<sup>-</sup> by CO<sub>2</sub>, decrease of pH, and conversion of CO<sub>2</sub> gas into inorganic carbon in growth medium, presented in Fig. 2. Chemically captured CO<sub>2</sub> can later be biologically converted into organic compounds by photosynthetic microorganisms (*S. platensis* in this study). The economically valuable products by photosynthetic microorganisms would offset the costs of CO<sub>2</sub> capturing absorbents and plant maintenance, although other costs such as isolating valuable product from remaining biomass mixtures have to be considered. CO<sub>2</sub> absorption capacity (kg CO<sub>2</sub>/kg absorbent) by several absorbents is presented in Table 1. From our experimental results, the chemical absorption by NaOH showed high CO<sub>2</sub> absorption capacity of 0.85 (kg CO<sub>2</sub>/kg NaOH). This CO<sub>2</sub> convergence process using NaOH for *S. platensis* cultivation has several advantages: fast chemical absorption of CO<sub>2</sub> gas,

reduction of cultivation costs by replacing NaHCO<sub>3</sub> with NaOH in standard Zarrouk medium for growth of *S. platensis*, biological conversion of fixed CO<sub>2</sub> into valuable compounds, and decreased sterilization costs using alkalinity of NaOH solution. Depending on the cultivation place of *S. Platensis*, there may be either a carbon tax or carbon credit, which provides the minus value of carbon dioxide in conjunction with economic analysis. In this study, it is assumed that the place of *S. Platensis* cultivation is located in Korea, which at the moment provides the carbon trading market at approximately US\$10 per CO<sub>2</sub> ton (Table 2). Based on the industry sources (<http://www.kosep.co.kr> and <http://kospo.co.kr>), the emitted gas from coal powered plant has the approximate composition of CO<sub>2</sub> 10.2%, O<sub>2</sub> 2.8%, and N<sub>2</sub> 87%. The emitted CO<sub>2</sub> from coal-powered plant can be directly applied for reaction with sodium hydroxide. Although a comprehensive analysis of including labor cost, *Spirulina* product price, raw material, interest rate, capital investment etc., will be considered in this work for the purpose of small-scale economic feasibility, we limited the economical viewpoint within the advantages due to substitution of nutrient related with CO<sub>2</sub> absorption. In conclusion, the emitted gas composition of coal-powered plant can be used directly on a commercial scale. This work presents not only the reduction in preparation cost of culture medium for *Spirulina* by 34.3% but also a distinct advantage involved with global warming.

Sterile conditions in a closed system, which is important for reliable mono-cultivation, can be usually achieved by heat- and chemical-treatment. Autoclave is the most reliable method to eliminate population of fungi, bacteria, viruses and spores, although it is energy-consuming and inappropriate for heat-sensitive materials such as plastics and electronic sensors. Chemical sterilization, widely used medically, avoids the problem of heat damage. However, its proce-

**Table 1. CO<sub>2</sub> absorption capacity by several chemical absorbents**

Absorbent	CO <sub>2</sub> absorption capacity (kg CO <sub>2</sub> /kg absorbent)	
NaOH	0.85	In this study
Monoethanol amine (MEA)	0.40	[17]
NH <sub>3</sub> (Aqua ammonia)	1.20	"
Dual alkali (MEA+NaCl)	0.54	"
Na <sub>2</sub> CO <sub>3</sub>	0.73	"
Aminoethylethanolamine (AEEA)	0.42	[7]
Piperazine (PZ)	0.51	"
Methyldiethanolamine (MDEA)	0.37	"
2-Amino-2-methyl-1-propanol (AMP)	0.49	"
Diethylenetriamine (DETA)	0.43	"

**Table 2. Cost advantage by CO<sub>2</sub> absorption at the preparation of culture medium**

	Concentration (g/L)	Cost (\$/kg) <sup>a</sup>
NaHCO <sub>3</sub>	16.80 (Zarrouk medium)	0.503 <sup>a</sup>
NaOH	8.00 (used amount in the study)	0.701 <sup>a</sup>
CO <sub>2</sub> (Carbon Tax, Korea)	6.79 (absorbed amount in the study)	-0.009 <sup>b</sup>

<sup>a</sup>ICIS.COM (2008) Indicative Chemical Prices A-Z, retrieved on August 20, 2014, from <http://www.icis.com/chemicals/channel-info-chemicals-a-z/>

<sup>b</sup>Estimated Carbon Emission Trading Price, Korea, <http://www.gir.go.kr>

dures require multiple washes with sterile water after treatment with sterilizing agents. These chemicals are often toxic and can be harmful to the environment. Therefore, a cost-efficient and eco-friendly sterilization process is necessary for validating the economic feasibility and achieving sustainable technology of microalgae-based biorefinery. Sodium hydroxide is widely used for cleaning and sanitization [19]. In the present study, incubation of culture medium with NaOH for 1 hours resulted in successful reduction of bacterial populations. High alkalinity induced by NaOH was neutralized to pH 8 or pH 9 by chemical absorption of CO<sub>2</sub> gas prior to cultivation (Fig. 2). The applied chemical absorption process using NaOH could avoid additional cost-consuming steps caused by unavoidable neutralization and washing steps when other chemical sterilants were used. This approach of sanitization can be applied to cultivations of other microalgae and cyanobacteria, which require large amount of bicarbonate for their growth. In this work, *S. platensis* served as a model organism.

### CONCLUSIONS

This work describes a sequential process involving chemical absorption of CO<sub>2</sub> gas and cultivation of *S. platensis*. Bicarbonate in standard Zarrouk medium could be completely replaced by inorganic carbonic ion, formed by chemical absorption of 154.2 mM CO<sub>2</sub> using 0.2 M NaOH. In conjunction with the carbon emission trading market in Korea, it is assumed that the cultivation place of *S. Platensis* is located in Korea and, accordingly, this process enabled us to reduce the preparation cost of Zarrouk medium by 34.3% via complete replacement of NaHCO<sub>3</sub> by NaOH (Table 2). Moreover, high alkalinity induced by NaOH solution could be used for cost-efficient and eco-friendly sterilization of culture medium. In this research, the use of inorganic carbon sources via chemical absorption of CO<sub>2</sub> does not decrease the amount of biomass and pigments in *S. platensis* compared to using standard Zarrouk medium.

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### REFERENCES

1. A. Belay, Y. Ota, K. Miyakawa and H. Shimamatsu, *J. Appl. Phycol.*, **5**, 235 (1993).
2. D. White, A. Pagarette, P. Rooks and S. Ali, *J. Appl. Phycol.*, **25**, 153 (2013).
3. S.-H. Ho, C.-Y. Chen, D.-J. Lee and J.-S. Chang, *Biotechnol. Adv.*, **29**, 189 (2011).
4. C. Stewart and M.-A. Hessami, *Energy Convers. Manage.*, **46**, 403 (2005).
5. C. González-López, F. Acien Fernández, J. Fernández-Sevilla, J. Sánchez Fernández and E. Molina Grima, *Biotechnol. Bioeng.*, **109**, 1637 (2012).
6. Y. Peng, B. Zhao and L. Li, *Energy Procedia*, **14**, 1515 (2012).
7. C.-H. Yu, C.-H. Huang and C.-S. Tan, *Aerosol Air Qual. Res.*, **12**, 745 (2012).
8. R. Moran, *Plant Physiology*, **69**, 1376 (1982).
9. A. Patel, S. Mishra, R. Pawar and P. Ghosh, *Protein Expression and Purification*, **40**, 248 (2005).
10. K. H. Ogbonda, R. E. Aminigo and G. O. Abu, *Bioresour. Technol.*, **98**, 2207 (2007).
11. A. Kumari, A. Kumar, A. K. Pathak and C. Guria, *J. CO<sub>2</sub> Util.*, **8**, 49 (2014).
12. M. Kanniche, R. Gros-Bonnivard, P. Jaud, J. Valle-Marcos, J.-M. Amann and C. Bouallou, *Appl. Therm. Eng.*, **30**, 53 (2010).
13. J. D. Figueroa, T. Fout, S. Plasynski, H. McIlvried and R. D. Srivastava, *Int. J. Greenhouse Gas Control*, **2**, 9 (2008).
14. R. Thiruvengatachari, S. Su, H. An and X. X. Yu, *Prog. Energy Combust. Sci.*, **35**, 438 (2009).
15. J. Pires, F. Martins, M. Alvim-Ferraz and M. Simões, *Chem. Eng. Res. Des.*, **89**, 1446 (2011).
16. D. W. Keith, M. Ha-Duong and J. K. Stolaroff, *Clim. Change*, **74**, 17 (2006).
17. B. P. Spigarelli and S. K. Kawatra, *J. CO<sub>2</sub> Util.*, **1**, 69 (2013).
18. O. Pulz and W. Gross, *Appl. Microbiol. Biotechnol.*, **65**, 635 (2004).
19. G. Healthcare, *Appl. Note*, **18**, 1124 (2009).